

# Evolution of Gigantism in Amphiumid Salamanders

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## Abstract

The Amphiumidae contains three species of elongate, permanently aquatic salamanders with four diminutive limbs that append one, two, or three toes. Two of the species, *Amphiuma means* and *A. tridactylum*, are among the largest salamanders in the world, reaching lengths of more than one meter, whereas the third species (*A. pholeter*), extinct amphiumids, and closely related salamander families are relatively small. *Amphiuma means* and *A. tridactylum* are widespread species and live in a wide range of lowland aquatic habitats on the Coastal Plain of the southeastern United States, whereas *A. pholeter* is restricted to very specialized organic muck habitats and is syntopic with *A. means*. Here we present analyses of sequences of mitochondrial and nuclear loci from across the distribution of the three taxa to assess lineage diversity, relationships, and relative timing of divergence in amphiumid salamanders. In addition we analyze the evolution of gigantism in the clade. Our analyses indicate three lineages that have diverged since the late Miocene, that correspond to the three currently recognized species, but the two gigantic species are not each other's closest relatives. Given that the most closely related salamander families and fossil amphiumids from the Upper Cretaceous and Paleocene are relatively small, our results suggest at least two extreme changes in body size within the Amphiumidae. Gigantic body size either evolved once as the ancestral condition of modern amphiumas, with a subsequent strong size reduction in *A. pholeter*, or gigantism independently evolved twice in the modern species, *A. means* and *A. tridactylum*. These patterns are concordant with differences in habitat breadth and range size among lineages, and have implications for reproductive isolation and diversification of amphiumid salamanders.

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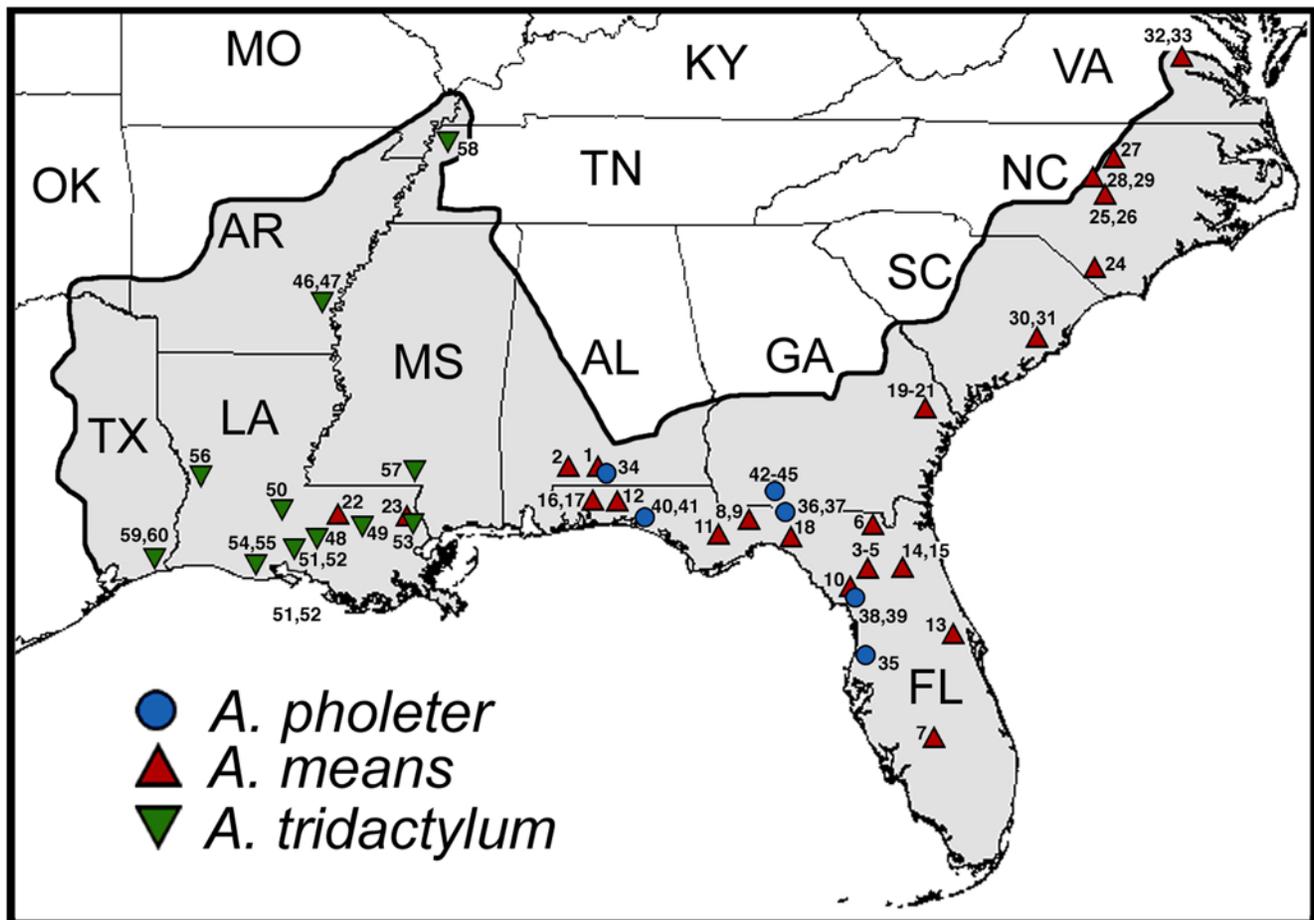
## Introduction

Body size evolution is a key factor in generating ecological and genetic divergence, and has been a primary axis of change during the radiation of many species groups. This is because body size is a relatively labile character, yet can be important for dictating niche parameters, creating reproductive isolation, and structuring communities [1–5]. Furthermore, physical attributes of an organism's size can influence other ecological parameters such as dispersal capabilities and habitat specialization [6–9]. Some groups of organisms have experienced extreme, and often paradoxical, evolutionary changes in body-size (gigantism and miniaturization) due to colonizing new regions such as islands or deep seas [10–13], re-colonization after mass extinctions [14], *in situ* environmental shifts [15–19], or evolutionary novelty [19]. Selection for gigantic body size is favored in situations of increased resource abundance, ecological release from predators or competitors, or necessity for long distance dispersal, whereas, miniaturization often results from resource or habitat limitations.

Salamanders of the family Amphiumidae inhabit lowland aquatic habitats throughout the Coastal Plain of the southeastern United States, and constitute an average 30 to 45 fold difference in body-size among species, including two of the largest salamanders in the world [20, this study]. Members of the genus *Amphiuma* are

elongate with expanded trunks and four miniature limbs, which is a major morphological deviation from the standard salamander body plan. The three currently recognized species are diagnosed by the number of toes on each limb: the one-toed (*Amphiuma pholeter*), two-toed (*A. means*), and three-toed (*A. tridactylum*) amphiuma. *Amphiuma means* and *A. tridactylum* are truly gigantic salamanders reaching lengths of more than a meter, whereas *A. pholeter*, the smallest species, reaches only ~36 cm in total length [20, this study]. *Amphiuma means* and *A. tridactylum* occur in a wide range of lowland aquatic habitats in the eastern and western parts of the Coastal Plain respectively (Figure 1), and may hybridize in the mid-Gulf Coastal Plain [21]. *Amphiuma pholeter* is restricted to organic muck habitats along the margin of the eastern Gulf Coastal Plain where it is microsympatric with juvenile *A. means*.

Analyzing the evolution of characteristics, such as body size, in a phylogenetic context can be fundamental to understanding the pattern and direction of change [22]. Previous molecular analyses of the Amphiumidae based on allozymes [23] and mitochondrial DNA sequences [24] found conflicting relationships, although both studies were based on limiting sampling. The discrepancies between these studies could be due to discordance between mitochondrial and nuclear gene phylogenies or sampling from different parts of the distribution (e.g., if the allozyme samples were



**Figure 1. Distribution and sampling of *Amphiuma*.** Map shows the combined distribution of the Amphiumidae (shaded in grey), and sampling localities for *A. pholeter* (blue circles), *A. means* (red triangles), and *A. tridactylum* (green triangles). Localities are listed in Table S1. doi:10.1371/journal.pone.0005615.g001

from zones of intergradation). Here we present sequence data for the mitochondrial genes cytochrome *b* (*Cytb*) and *16 s* and a nuclear recombination activating gene-1 (*Rag1*) for *Amphiuma* from across the geographic distribution of the family to estimate lineage diversity. To further reconstruct the relationships among these lineages, we used ~4 Kb of mitochondrial and ~3 Kb of nuclear DNA. Using this robust phylogeny for the family and considering ancient fossil amphiumids, we present hypotheses for the evolution of extreme body size changes and its implications for ecological and genetic divergence in this family.

## Methods

### Sampling

Sixty *Amphiuma* tissue samples were collected from across the distribution of the three currently recognized species or obtained from museum collections (Figure 1, Table S1). The species were preliminarily identified by a combination of their geographic distribution, toe number, and body coloration. All specimens were handled in accordance with Institutional Animal Care and Use Committee (IACUC) protocols at the University of Tulsa, University of California, Berkeley, and the University of Texas at Arlington.

### DNA sequence collection and alignment

DNA was isolated from fresh frozen or ethanol preserved tissues using Qiagen DNeasy extraction kits. To estimate lineage diversity

of *Amphiuma* we amplified portions of two mitochondrial genes, *Cytb* (783 bp) and *16 s* (538 bp), and the nuclear gene *Rag1* (825 bp) from specimens from across the distribution of the three species (Figure 1; Table S1). To further test relationships among the major lineages of *Amphiuma*, we amplified portions of the mitochondrial genes cytochrome oxidase-1 (*CoI*, 1260 bp) and NADH dehydrogenase subunit 4 (*Nd4*) and adjacent tRNAs (886 bp), and the nuclear genes *Rag1* (1525 bp), pro-opiomelanocortin (*Pomc*, 481 bp), sodium-calcium exchanger 1 (*Ncx1*, 814 bp), and solute carrier family 8 member 3 (*Slc8a3*, 761 bp). For phylogenetic analyses we used outgroups from three other salamander families: *Ambystoma mexicanum* (Ambystomatidae), *Plethodon cinereus* (Plethodontidae), and *Rhyacotriton variegatus* (Rhyacotritonidae). Most outgroup sequences were taken from Genbank (Tables S1, S2; [25–31]), whereas others were collected using the methods described below. The PCR primers used are listed in Table S3 [32–35].

PCR products were run on 1% agarose gels, and products of the expected molecular weight were cleaned with either a Millipore PCR<sub>96</sub> cleanup kit (Montage™) or ExosapIT (USB Corp). Cycle sequencing reactions using Big Dye v. 3.1 (Applied Biosystems Inc.) were cleaned with Sephadex (Sigma) and sequenced on either an ABI 3730 or 3130xl capillary sequencer. Individual sequences were edited and translated in Sequencher™ vers 4.8 (Gene Codes Corp). The alignments of protein coding genes were unambiguous, but some length variable regions of the ribosomal

gene *16s* and the tRNAs that were questionable were removed prior to analyses. The lengths of alignments were also trimmed so all sequences in a given alignment were the same length. The final alignment of the geographic variation datasets includes 60 individuals of mtDNA from *Cytb* (651 bp) and *16s* (377 bp) and 50 individuals of nuclear DNA from *Rag1* (548 bp). For the combined analysis of representative lineages we used a total of 2,948 bp of nuclear DNA from: *Rag1* (1,474 bp), *Pomc* (481 bp), *Ncx1* (735 bp) and *Slc8a3* (258 bp). We also used 4,068 bp of mtDNA from the genes: *Cytb* (779 bp), *16s* (377 bp), *Col1* (517 bp), *Nd4* (629 bp), *tRNA<sup>HIS</sup>* (54 bp), and also NADH dehydrogenase subunits 1 (*Nd1*, 335 bp) and 2 (*Nd2*, 1,010 bp), and adjacent tRNAs *<sup>ILE</sup>*, *tRNA<sup>MET</sup>*, *tRNA<sup>TRP</sup>*, *tRNA<sup>ALA</sup>*, *tRNA<sup>ASN</sup>*, and *tRNA<sup>ILE</sup>* (367 bp total) from Genbank [24,26,29].

### Phylogenetic analyses and divergence time estimates

Individual genes and datasets including combinations of genes were analyzed with Maximum Parsimony (MP) and Bayesian (BA) methods. Unweighted maximum-parsimony analyses using heuristic searches with 100 random-taxon-addition replicates, and non-parametric bootstrapping [36] based on 1000 pseudoreplicates and 10 random taxon-addition-replicates per pseudoreplicate, were performed in PAUP\* v. 4.0b10 [37]. MrModeltest v. 2.2 [38] was used to determine the most appropriate model of nucleotide substitution for each data partition (Table S4). The datasets were partitioned by gene, and protein coding genes were further partitioned by codon position [28,39,40]. Alignments including primarily evolutionarily conserved stems of seven tRNAs were combined and analyzed under a single model. Partitioned Bayesian analyses (all partitions unlinked) implemented via MrBayes v. 3.1 [41,42] were run with four chains (three hot and one cold) and uniform priors for five million generations (with a tree saved at every 1000 generations). We discarded the first one million generations (1000 trees) as burn-in. The resulting 50% majority-rule consensus of the 4001 post burn-in trees, sampled every 100 generations, was computed in PAUP\* [37]. By default, MrBayes 3.1 runs each analysis twice simultaneously, and in each case our independent runs converged on the same topology and posterior probabilities for all of the major nodes. We used the Shimodaira-Hasegawa test (SH-test; [43]) implemented in PAUP\* [37] to specifically test among the three alternate hypotheses for the relationships of amphiuma: 1. (*A. pholeter* (*A. means*+*A. tridactylum*)); 2. (*A. means* (*A. pholeter*+*A. tridactylum*)); and 3. (*A. tridactylum* (*A. means*+*A. pholeter*)). The SH-test was based on the complete 7 Kb dataset analyzed using GTR+ $\Gamma$  and base frequencies and rate matrix determined by MrModeltest [38].

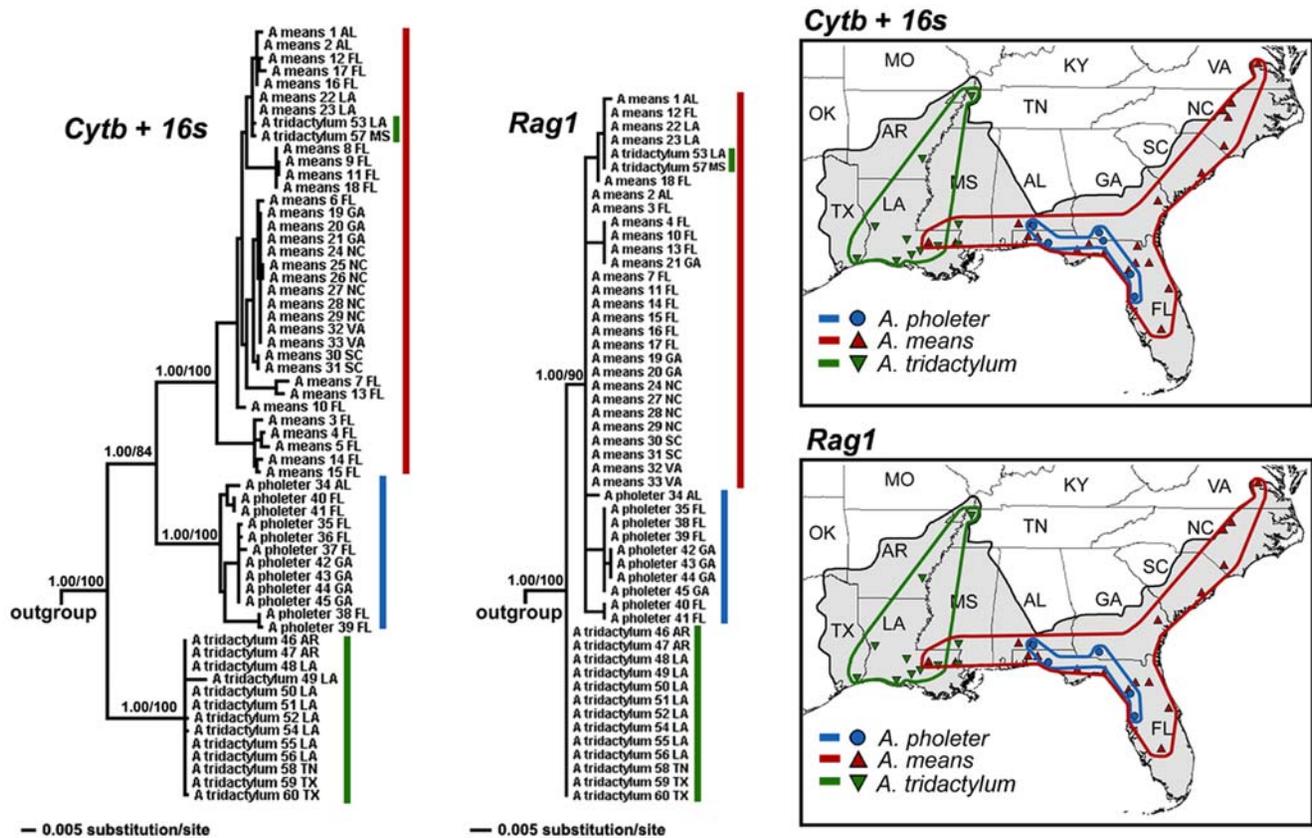
We estimated divergence times using penalized likelihood (PL) in the program r8s v. 1.7 [44,45], based on a *Rag1* family-level phylogeny of salamanders with the topology and branch lengths estimated via a partitioned Bayesian analysis in MrBayes [41,42] (Table S5). The Bayesian analysis of *Rag1* (1,410 bp) was run using four chains (3 hot and 1 cold) for five million generations with a tree saved every 10,000 generations. The first 100 trees (one million generations) were discarded as burnin and the 400 post burnin trees were used to estimate the topology and branch lengths used to estimate the ages of select nodes. The tree was rooted with a caecilian (*Ichthyophis*) and a frog (*Ascaphus montanus*) was also included as an outgroup for the phylogenetic analysis, but both taxa were pruned in r8s prior to calculating divergence times. We fixed the basal split between two major lineages of crown group salamanders (cryptobranchoids and salamandroids) at two different dates: 1) 161 MY, based on the earliest known cryptobranchoid, *Chunerpeton taiyuiensis* [46], and 2) 250 MY, which is an approximate average between some of the oldest

molecular based divergences for this split which range from 220 to 275 MYA [30,31,47]. We also used four fossil salamandroids to serve as minimum external calibration points (Table S6; Figures S1, S2 [48–52]). The TN (truncated Newtonian) method was used for PL, and the cross validation procedure was run in eight increments of 0.5 from 0 to 3.5 (on a log10 scale) to test for the optimal smoothing parameter for analyses with the basal node fixed at either 161 or 250 MYA. The optimal smoothing parameters were 32 (161 MY) and 100 (250 MYA). The profile command was used to calculate the mean age and standard deviation for select nodes based on the branch lengths of the 400 post burnin Bayesian trees.

### Analysis of body-size evolution

To assess the extent of body size differences among modern *Amphiuma*, we measured the length and girth of adults from museum collections. Body length was based on measuring both total length (TL = tip of snout to the tip of tail) and snout to vent length (SVL = tip of the snout to the posterior margin of the cloaca), and girth was estimated by measuring the body depth (BD) and body width (BW) immediately anterior to the forelimbs. All specimens measured were at or above the minimum adult body sizes reported for each of the three species: *A. tridactylum*, 33 cm SVL [53]; *A. means*, 26 cm SVL [54]; and *A. pholeter*, 19 cm SVL (based on 24 cm TL [55]). *Amphiuma* are relatively cylindrical in shape, so we estimate the average overall body size (head and trunk) for each species by calculating body volume using the formula for an elliptical cylinder =  $\pi \times (\text{major axis}/2) \times (\text{minor axis}/2) \times \text{Length}$ , where the major axis = BW, minor axis = BD, and length = SVL. Maximum total lengths reported in the literature for these species are: *A. tridactylum*, 106 cm [56], *A. means*, 116 cm [56,57], and *A. pholeter*, 33 cm [55]. Body lengths of fossil amphiumids and outgroups were taken or estimated from the literature. The small isolated vertebrae of *Proamphiuma cretacea* have well developed crests and heavy ossification, so they are presumed to be from adults estimated to be ~30 cm TL [49]. Specimens assignable to *Amphiuma jepseni* are limited, but based on its description [48] and the size of the vertebrae, we infer that this specimen is also an adult of small size (~30 cm TL). Several recent higher-level studies of salamander phylogeny support a clade that includes the families Rhyacotritonidae, Plethodontidae, and Amphiumidae, with strong support for a sister relationship between amphiumids and plethodontids [27,28,30]. Rhyacotritonids are not known from the fossil record, and all four extant species are small (adults 7 to 11.5 cm TL; [58]). Similarly, most plethodontid genera comprise relatively small species, and the few “large” species are no longer that *A. pholeter* [20,59]. Therefore, we consider *A. means* and *A. tridactylum* gigantic species, as they are among the largest extant amphibians, and based on the information above we consider *P. cretacea*, *A. jepseni*, *A. pholeter*, plethodontids, and rhyacotritonids to be small taxa.

Mesquite v 2.5 [60] was used to analyze the ancestral states of the trait maximum body size (TL) as a discrete character (gigantic vs small) using a likelihood framework. This method allows the rate of change between states to be modeled when tracing the evolution of characters on the phylogenetic tree, and calculates the proportional likelihood of the ancestral condition for each node. Our analyses were based on the Markov k-state 1 (Mk1) parameter model that considers an equal rate of change between states. Reconstructions were based on the topology of the 7 Kb molecular dataset with the fossil taxa included based on their taxonomy and distribution in the fossil record (*P. cretacea* (*A. jepseni*+clade based on our estimate of the relationships among extant *Amphiuma*)). One advantage of Maximum Likelihood



**Figure 2. Geographic genetic variation in the Amphiumidae.** Bayesian phylograms of the mitochondrial genes *Cytb* and *16s* (left) and the nuclear gene *Rag1* (right). Numbers subtending the major nodes are Bayesian posterior probabilities to the left of the slash and maximum parsimony bootstrap values to the right. The three major clades, which primarily correspond to the recognized species, are indicated on the phylograms and maps to the right with colored lines: *A. pholeter* (blue), *A. means* (red), and *A. tridactylum* (green). Two putative "*A. tridactylum*" that are closely related to some *A. means* are highlighted on the phylogenies with green triangles.  
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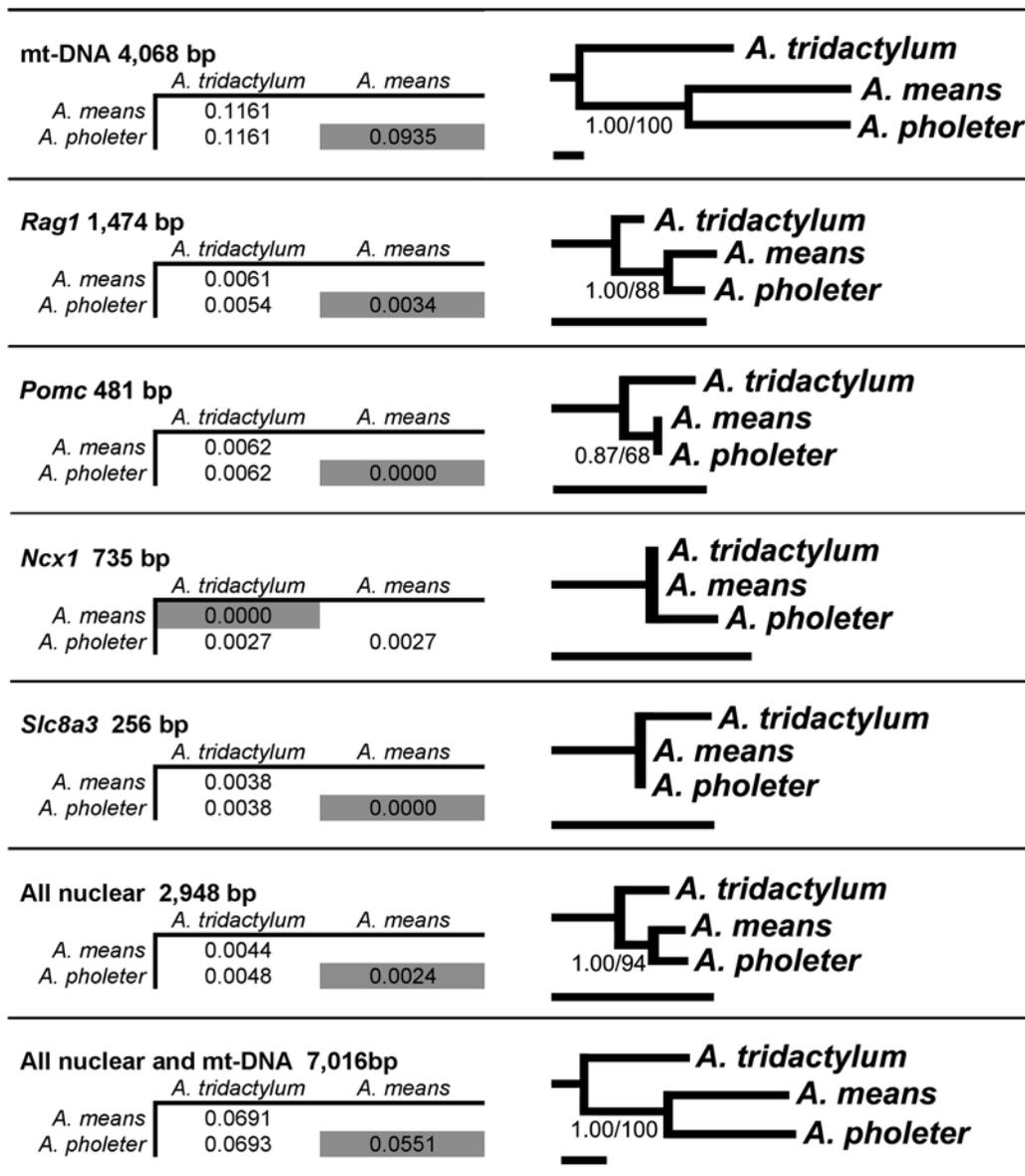
reconstruction of ancestral states (over parsimony based methods) is the ability to incorporate a time component that is estimated by branch lengths on the phylogeny. We calculated the proportional likelihood of the ancestral condition with and without branch lengths from Bayesian analysis of *Rag1*.

## Results

Geographic variation of mitochondrial DNA (based on *Cytb* and *16s*) indicates that modern amphiumas contain three divergent genetic lineages that nearly exactly correspond to the three recognized species (Figure 2). Herein we will refer to these lineages with their current taxonomic names. We found almost no variation in mtDNA within *A. tridactylum* (uncorrected  $P < 0.1\%$ ) from throughout their distribution. Some variation exists within *A. means* (uncorrected  $P = 4.37\%$ ), primarily within the Gulf Coastal Plain and Florida; populations across the entire Atlantic Coastal Plain from Georgia to Virginia are very similar (uncorrected  $P < 0.5\%$ ). We found mitochondrial variation of up to 2.5% among *A. pholeter* from different river drainages. We found little genetic variation in *Rag1* from across the distribution of the three species. However, even with a low level of variation at this locus, *A. means* and *A. pholeter* form a clade, primarily exclusive of *A. tridactylum*. We found no variation in *A. tridactylum* for *Rag1*. There are some nucleotide substitutions among *A. means* and *A. pholeter*, and these species do not form reciprocally monophyletic clades, which may

result from incomplete lineage sorting or too little variation to build an accurate tree. We did not find any cases where *A. means* and *A. pholeter* had identical *Rag1* sequences. Two specimens initially identified as *A. tridactylum* (53 and 57) from the Pearl River drainage were identical to *A. means* from the same drainage in both mtDNA and *Rag1*. However, we reexamined specimen 53 and found that it has two toes on some limbs and three toes on others, but it was not heterozygous for any of the otherwise diagnostic nucleotide differences between *A. means* and *A. tridactylum*. Specimen 57 is not available for morphological reexamination, but it had three toes on at least some limbs when collected. We interpret these specimens to be either hybrid backcrosses between *A. means* and *A. tridactylum*, or simply *A. means* with anomalous numbers of toes on some limbs, perhaps a recurrence of the ancestral condition.

The uncorrected pairwise divergence of mitochondrial genes and three of the four nuclear loci show *A. tridactylum* to be the most divergent lineage of *Amphiuma*, whereas *A. means* and *A. pholeter* are most similar (Figure 3). Only the nuclear gene *Ncx1* shows *A. pholeter* to be the most divergent lineage of *Amphiuma*, but that is based upon only a single substitution in *A. pholeter* out of 735 bp; the three species are otherwise identical. Phylogenetic analyses show strong support for *A. means* and *A. pholeter* as a clade exclusive of *A. tridactylum*, based on mitochondrial DNA alone (BAPP = 1.00, MPBS = 100), the combined nuclear data (BAPP = 1.00, MPBS = 94), and the combined mitochondrial and nuclear data



**Figure 3. Genetic divergence and phylogenetic relationships for the nominate taxa of *Amphiuma* based on mitochondrial and nuclear genes.** Matrices to the left are uncorrected pairwise sequence divergence among the three taxa with the most closely related pair highlighted in grey. Bayesian phylograms on the right describe the relationships among the three taxa, and statistical support, if any, is indicated by Bayesian posterior probabilities to the left of the slash and maximum parsimony bootstrap values to the right.  
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(BAPP = 1.00, MPBS = 100). Analyses of each nuclear gene alone support either the *A. means*+*A. pholeter* clade (*Rag1* and *Pomc*) or were unable to resolve the relationships among the lineages (*Ncx1* and *Slc8a3*). Maximum likelihood analyses based on the combined dataset provided strong support for an *A. means*+*A. pholeter* clade (MLBS = 100). Furthermore, SH-tests show this topology to be significantly more likely than the ML phylogeny with *A. means* and *A. tridactylum*, or *A. pholeter* and *A. tridactylum*, constrained to be monophyletic (Table 1). Means and Karlin's [23] genetically similar samples of *A. means* and *A. tridactylum* fall well within our genetically divergent geographic lineages of *A. means* and *A. tridactylum* for both mt-DNA and *Rag1*. Although we find very low levels of nuclear variation in *Amphiuma*, it is peculiar that they found *A. pholeter* to be so divergent from *A. means* and *A. tridactylum*. The only way that we can reconcile this discrepancy is if all of

Means and Karlin's [23] allozyme loci are geographically discordant with *Rag1* and mitochondrial variation, or if they have had increased rates of evolution in the *A. pholeter* lineage.

Divergence time estimates based on penalized likelihood of *Rag1* show a most recent common ancestor of modern *Amphiuma* (i.e., the split between *A. tridactylum* and *A. means*+*A. pholeter*) to be  $5.0 \pm 1.5$  MYA or  $7.8 \pm 2.3$  MYA, and divergence between *A. means* and *A. pholeter* to be  $2.1 \pm 0.8$  MYA or  $3.2 \pm 1.2$  MYA. These alternate dates for each node represent analyses based on fixing the basal node (cryptobranchoids+salamandroids) at either 161 MYA [46] or 250 MYA [30,31,47] respectively (See Methods; Tables 2, S6; Figures S1, S2).

Our body size and girth measurements show that average adult *A. means* and *A. tridactylum* are about 2.5 times longer, and ~3.5 to 4 times wider and deeper than *A. pholeter* (Table 3; Figure 4). Taken

**Table 1.** Results of Shimodaira-Hasegawa tests of two constrained alternate topologies to an unconstrained maximum likelihood analysis based on the combined mitochondrial and nuclear dataset.

test	topology	-ln likelihood	Difference in -ln likelihood	p
	Unconstrained <i>A. means</i> and <i>A. pholeter</i> monophyletic	24811.97	---	---
1	<i>A. means</i> and <i>A. tridactylum</i> monophyletic	24845.88	33.91	<0.001
2	<i>A. pholeter</i> and <i>A. tridactylum</i> monophyletic	24847.29	35.32	<0.001

doi:10.1371/journal.pone.0005615.t001

**Table 2.** Results of divergence time estimates (Average±Standard deviation) based on a Bayesian *Rag1* phylogeny of salamanders.

Node	PL (161)	PL (250)
<i>A. means</i> + <i>A. pholeter</i>	2.1±0.8	3.2±1.2
<i>A. tridactylum</i> + <i>A. means</i> + <i>A. pholeter</i>	5.0±1.5	7.8±2.3
Plethodontidae+Amphiumidae	78.1±7.6	121.8±12.0
Rhyacotritonidae+Plethodontidae+Amphiumidae	98.4±9.1	153.0±14.2

Dates were estimated using penalized likelihood (PL) and fixing the basal split between cryptobranchoids and salamandroids at either 161 MY or 250 MY.  
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together the gigantic species are on average >30 (*A. means*) and 45 (*A. tridactylum*) times larger (in volume) than *A. pholeter* (Table 3). This difference is the same whether we estimate overall body size (head and trunk) as an elliptical cylinder ( $\pi \times (BW/2) \times (BD/2) \times SVL$ ) or as a rectangular prism ( $BW \times BD \times SVL$ ). Maximum likelihood reconstruction of ancestral body size as a discrete character (gigantic vs small), assuming an equal rate of change between states, and considering extant and fossil taxa shows a marginally higher proportional likelihood for gigantism as the ancestral condition for modern *Amphiuma* (0.530) and also the ancestor of the clade *A. means*+*A. pholeter* (0.524; Table 4, Figure 5). The proportional likelihood for gigantism increases slightly for both modern *Amphiuma* (0.576) and also the ancestor of the clade *A. means*+*A. pholeter* (0.567) when *Rag1* branch length information is incorporated in the calculation (Table 4).

## Discussion

### Phylogeny and evolution of body-size in the Amphiumidae

The earliest molecular systematic study of all species of *Amphiuma*, based on allozymes, found the gigantic species, *A. means*

and *A. tridactylum*, to be closely related (Nei's  $D = 0.12$ ), whereas, *A. pholeter* is very divergent (Nei's  $D = 0.90$  from *A. means* and 0.73 from *A. tridactylum*) and represented an "ancient evolutionary offshoot" [23]. A more recent phylogenetic analysis of salamander families based on mitochondrial DNA sequences included an individual of all three species of *Amphiuma* and showed *A. means* and *A. pholeter* to be sister taxa [24]. Our analysis based on widespread sampling of both nuclear and mitochondrial DNA sequences across the geographic distribution of the Amphiumidae, reveals three primary genetic lineages that correspond to the three recognized species (Figure 2). Consistent with the second study [24], we found strong support for a sister relationship between *A. means* and *A. pholeter* (Figure 3, Table 1). Furthermore, our divergence time estimates indicate that *A. tridactylum* represents the earliest diverged lineage among modern species, whereas *A. means* and *A. pholeter* share a more recent common ancestor. We estimate the oldest divergence among modern lineages of *Amphiuma* to be no older than the Miocene, suggesting that the two definitive fossil amphiumids from the Upper Cretaceous (*Proamphiuma cretacea*) and Paleocene (*Amphiuma jepseni*) [52,53] are indeed outgroups. In summary our hypothesis for the relationships of the family Amphiumidae are: (*P. cretacea* (*A. jepseni* (*A. tridactylum* (*A. means*+*A. pholeter*))))).

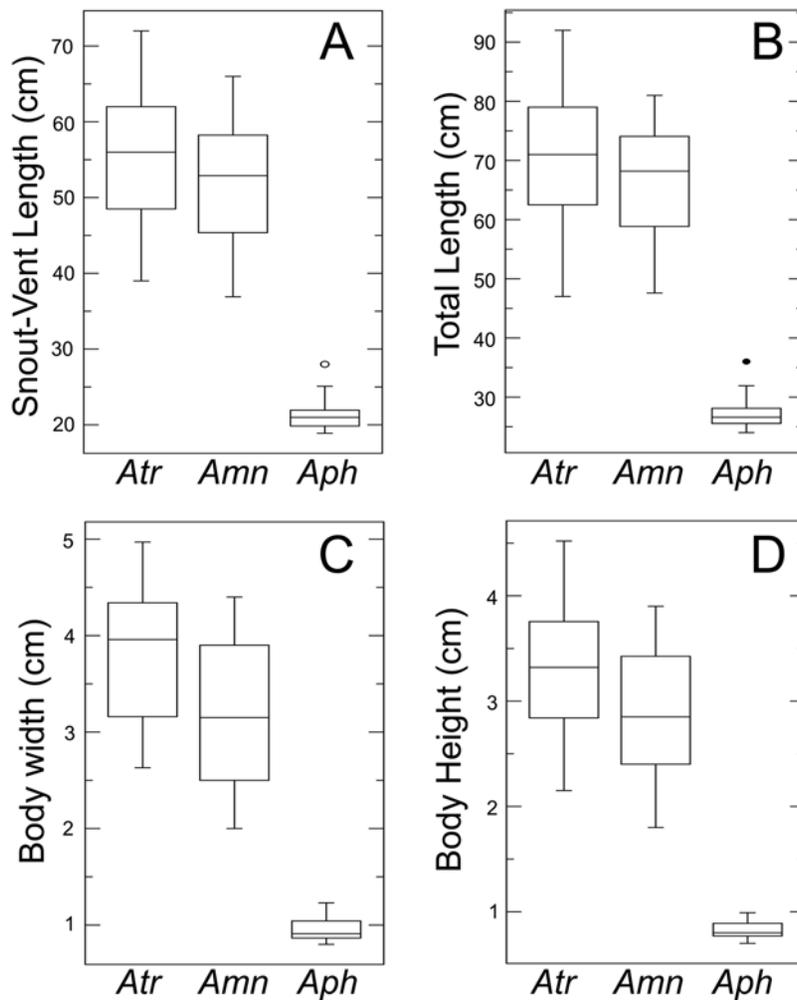
Closely related families of salamanders (rhyacotritonids and plethodontids), fossil amphiumids from the Upper Cretaceous (*Proamphiuma cretacea*) and Paleocene (*Amphiuma jepseni*), and *A. pholeter* are small, whereas *A. means* and *A. tridactylum* are gigantic (>30 to 45 times larger than *A. pholeter*). Therefore, our phylogeny and reconstruction suggest two possible scenarios for the evolution of gigantism in this family: 1) Gigantic body size either evolved once, since the Paleocene, and was the ancestral condition of modern amphiumas, with a subsequent strong size reduction in *A. pholeter* or 2) small body size was the ancestral condition of extant *Amphiuma* and gigantism independently evolved in the lineages leading to the two modern species *A. tridactylum* and *A. means*. Maximum likelihood reconstruction shows a marginally higher proportional likelihood for gigantism as the ancestral condition for modern *Amphiuma* and also for the most recent common ancestor

**Table 3.** *Amphiuma* body-size.

Species	n	SVL	TL	BW	BD	Head and trunk volume $\pi \times (BW/2) \times (BD/2) \times SVL$
<i>A. tridactylum</i>	25	55.4 (39.0–72.0)	70.5 (47.0–92.0)	3.8 (2.6–5.0)	3.3 (2.2–4.5)	545.6 (195.3–1128.6)
<i>A. means</i>	18	52.0 (36.9–66.0)	66.6 (47.6–81.0)	3.2 (2.0–4.4)	2.9 (1.8–3.9)	379.0 (99.3–812.2)
<i>A. pholeter</i>	12	21.4 (18.9–28.0)	27.4 (24.0–36.0)	0.9 (0.8–1.2)	0.8 (0.7–1.0)	12.1 (9.6–25)

Numbers in parenthesis represent upper and lower values for each measurement and the number above is the average. Measurements are in centimeters (cm) and volume estimate is in  $cm^3$ .

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**Figure 4. Body sizes of adult *Amphiuma*.** Box plots of A) snout-vent length, B) total length, C) body width, and D) body height for *A. tridactylum* (Atr), *A. means* (Amn), and *A. pholeter* (Aph) measured in this study. The five horizontal lines of each plot represent the minimum, first quartile, median, third quartile, and maximum values for each species. Filled circles represent outliers and open circles represent suspected outliers. Averages, and ranges for each measurement and species are listed in Table 3. doi:10.1371/journal.pone.0005615.g004

of *A. means* and *A. pholeter* (Figure 5). Given that our analysis of widespread geographic genetic variation revealed only three genetic lineages of modern amphiumids, our ability of further address the evolution of body-size by examining modern species is limited. However, the discovery of additional fossil lineages would greatly enhance our understanding of the evolution of body size of amphiumid salamanders. *Amphiuma antica* [61] was described from the mid-Miocene of Texas based on a single large, poorly preserved vertebra, but the assignment of this specimen to the Amphiumidae is questionable [49]. Gardner [49] also suggested that it could be one of the modern species of *Amphiuma*. We estimate that modern *Amphiuma* share a common ancestor in the late Miocene. If *A. antica* is gigantic *Amphiuma* and a stem or sister taxon to modern species, then this would strongly support our first hypothesis that the ancestor of modern *Amphiuma* was gigantic and the relatively small *A. pholeter* results from miniaturization. Regardless of the direction, or the number of times body size has changed in the Amphiumidae, this extreme change has happened over a relatively short period of time, since the sister taxa, *A. means* and *A. pholeter*, shared a common ancestor as recently as the late-Pliocene.

#### Niche breadth, distribution size, and reproductive isolation

Body size is a key parameter for determining the relative placement of an organism in its environment and can also impact its niche breadth, dispersal ability, and consequently, geographic distribution [6–9]. The extreme difference in body size among modern *Amphiuma* is coincident with strong differences in niche breadth and geographic distribution. The gigantic species, *A. means* and *A. tridactylum*, are widespread species that occur in diverse lowland aquatic environments, including swamps, ponds, marshes, rivers, and drainage ditches [20,62,63]. In contrast, the small species, *A. pholeter* has a relatively limited distribution and is restricted to specific organic muck (fine mud) habitats [55,64]. Our first scenario for the evolution of body size in the Amphiumidae suggests that a habitat specialist evolved via miniaturization of a gigantic, habitat generalist. The organic muck habitats where *A. pholeter* occur are derived from finely decayed plant matter that builds up as deep beds in lowland aquatic habitats. Interestingly, only juvenile *A. means* have been found syntopically with *A. pholeter* in the muck habitats [64; RWV and PEM, pers. obs.], so this unique habitat may only be favorable for small *Amphiuma*.

**Table 4.** Results of maximum likelihood ancestral state reconstruction of body-size, of extant *Amphiuma*, fossil amphiumids, and closely related families.

Node description	Likelihood Small	Likelihood Gigantic
<b>Without Branch lengths</b>		
<i>A. means</i> + <i>A. pholeter</i>	0.476	0.524
<i>A. tridactylum</i> + <i>A. means</i> + <i>A. pholeter</i>	0.470	0.530
<i>Amphiuma</i>	0.939	0.061
Amphiumidae	0.991	0.009
Plethodontidae+Amphiumidae	0.996	0.004
Rhyacotritonidae+Plethodontidae+Amphiumidae	0.985	0.015
<b>With Branch lengths</b>		
<i>A. means</i> + <i>A. pholeter</i>	0.433	0.567
<i>A. tridactylum</i> + <i>A. means</i> + <i>A. pholeter</i>	0.424	0.576
<i>Amphiuma</i>	0.987	0.013
Amphiumidae	0.997	0.003
Plethodontidae+Amphiumidae	0.959	0.041
Rhyacotritonidae+Plethodontidae+Amphiumidae	0.936	0.061

Analyses were run with and without Bayesian branch lengths. Both analyses were based on Mk1 model that assumes an equal rate of transition between the two states (small vs. gigantic).

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The estimated age for sexual maturity for *A. means* is 3 to 4 years (26 cm SVL) [65], whereas for *A. pholeter* it is only 2 years (19 cm SVL) or less [20]. Therefore, the miniature body size of *A. pholeter* may have occurred by early maturation (of a gigantic ancestor) during the evolution of a completely muck-dwelling existence. This semi-liquid muck appears to provide a substantial amount of support to the bodies of *A. pholeter*, as their locomotor ability is relatively limited in open water (RWV and PEM, pers. obs.). Even though fine muck habitats occur throughout the Coastal Plain, the relatively restricted current distribution of *A. pholeter* may result from geographic barriers, such as ridges and large rivers that limit their dispersal between muck habitats.

If scenario two is correct and recent ancestors of modern *Amphiuma* were small, two independent instances of gigantism are likely in the *A. means* and *A. tridactylum* lineages. This scenario implies parallel instances of Cope's Rule (evolutionary increase in body-size), where delaying maturation and drastically increasing overall body size would have had strong fitness consequences such as fecundity and survival. The Coastal Plain of the southeastern United States includes a wide range of lowland aquatic habitats. Gigantic body size may further allow *A. means* and *A. tridactylum* to traverse and colonize the wide breadth of habitats that occur across the Coastal Plain.

Evolution of body size is a simple mechanism for generating ecological and genetic divergence [1–5]. The shift in habitat use appears to be a distinct partition between gigantic and small *Amphiuma*, because *A. pholeter* spends almost its entire life in a specialized habitat that is not commonly utilized by the adults of *A. means*. This shift in habitat and body size may have provided a strong barrier for promoting genetic divergence between these species. Previous morphological analysis of the two gigantic species, *A. means* and *A. tridactylum*, from across their zone of overlap in the mid-Gulf Coastal Plain found them to be distinct species, but identified putative hybrid individuals from the Pearl River drainage that had an amalgam of otherwise species specific

traits, including specimens with two toes on some limbs and three toes on others [21]. Our samples from this region all had mitochondrial haplotypes and *Rag1* alleles similar to those of *A. means*, even though some had three toes on some limbs. In contrast, the distribution of the small species, *A. pholeter*, is entirely within that of one of the gigantic species, *A. means*. Despite the fact that they are the most closely related species of modern *Amphiuma*, these species, so strongly divergent in size, are not known to interbreed. More detailed genetic sampling along the contact zone is necessary to further test whether *A. means* and *A. tridactylum* hybridize. Also, more detailed sampling of microsympatric populations of *A. means* and *A. pholeter* would test if there has been any recent genetic interaction. If *A. means* and *A. tridactylum* interbreed but *A. means* and *A. pholeter* do not, body size and habitat specialization may serve as a greater isolating mechanism than genetic divergence alone.

In conclusion, our phylogeographic analyses based on both mitochondrial and nuclear DNA indicate three divergent lineages of modern *Amphiuma* that closely correspond to the three currently recognized species. Nearly all molecular data support a sister relationship between *A. means* and *A. pholeter* which diverged as recently as the late-Pliocene. When placing this relationship in the context of fossil amphiumids and closely related families we find that there has been either: 1) a single case of gigantism in the common ancestor of modern *Amphiuma* and subsequently a recent instance of miniaturization in *A. pholeter*, possibly as a specialization to a completely muck-dwelling existence, or 2) two independent instances of gigantism in *A. means* and *A. tridactylum*, which may contribute to their ability to traverse and inhabit a wide variety of lowland aquatic habitats. Additional fossils of mid-Cenozoic amphiumids will greatly enhance our understanding of the direction of body size evolution in these salamanders.

## Supporting Information

**Table S1** Specimen information and Genbank numbers 16 s, Cytb, and *Rag1* from across the distribution of all three species of *Amphiuma*.

Found at: doi:10.1371/journal.pone.0005615.s001 (0.07 MB DOC)

**Table S2** Specimen information and Genbank numbers for *Amphiuma* and outgroups used for individual and combined analyses of mitochondrial and nuclear loci (Figure 3). 16 s, Cytb, and *Rag1* for these analyses are listed in Table S1.

Found at: doi:10.1371/journal.pone.0005615.s002 (0.04 MB DOC)

**Table S3** Primers used for PCR and sequencing.

Found at: doi:10.1371/journal.pone.0005615.s003 (0.05 MB DOC)

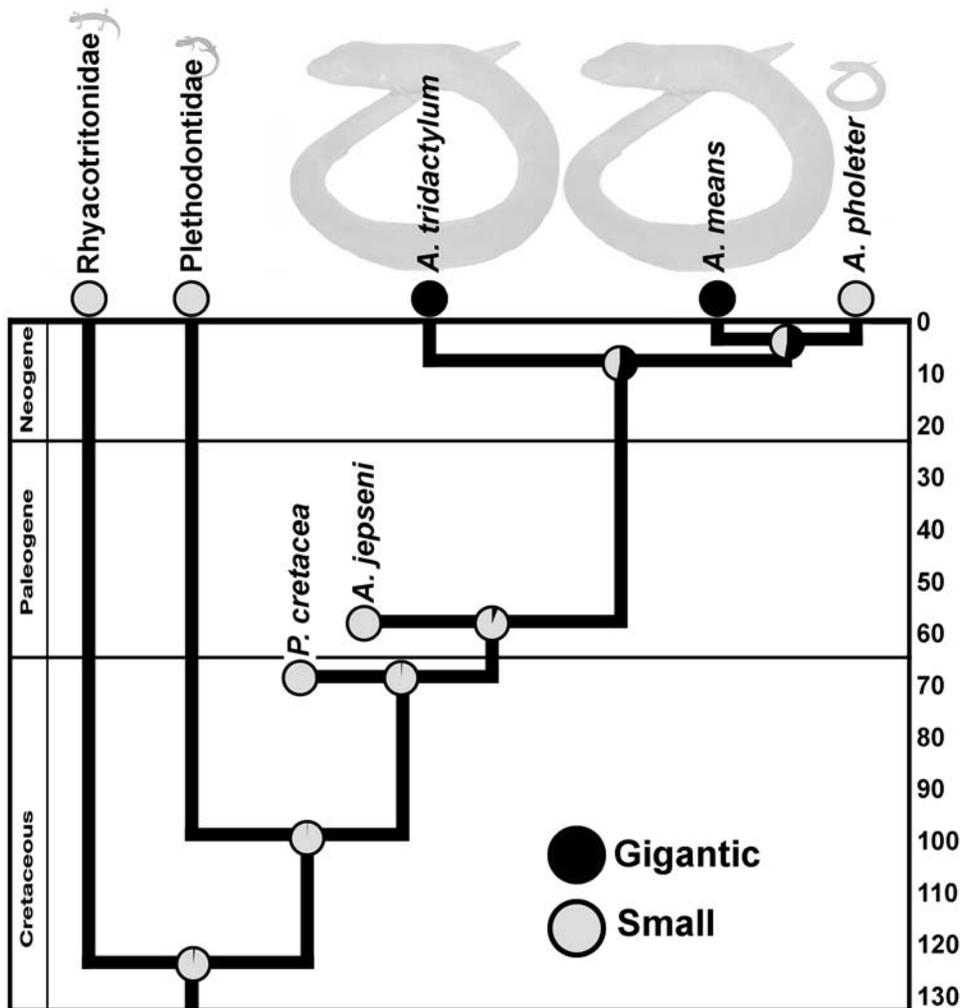
**Table S4** Models applied to each data partition for Bayesian analyses.

Found at: doi:10.1371/journal.pone.0005615.s004 (0.05 MB DOC)

**Table S5** Species and Genbank numbers for Bayesian phylogenetic analysis of *Rag1* that was used for divergence time estimates in r8s.

Found at: doi:10.1371/journal.pone.0005615.s005 (0.04 MB DOC)

**Table S6** External calibration points used for nonparametric rate smoothing analysis of *Rag1* using r8s. Points are plotted on Figures S1 and S2.



**Figure 5. Divergence time estimates and body-size evolution of amphiumid salamanders.** Extant taxa on the chronogram are drawn half way between the results of the two different penalized likelihood analysis of *Rag1* from r8s (Table 2). Extinct amphiumids were added to the tree based on their distribution in the fossil record and suggested relationships [49]. Reconstruction of ancestral body size (gigantic vs small) was performed using a maximum likelihood algorithm in Mesquite. Pie diagrams at each node indicate the likelihood of each state. The ancestral state data shown are based on analyses without branch lengths (Table 4). doi:10.1371/journal.pone.0005615.g005

Found at: doi:10.1371/journal.pone.0005615.s006 (0.04 MB DOC)

**Figure S1** Chronogram of salamander families calculated in r8s based on Bayesian analysis of *Rag1*, fixing the basal node at 161 MYA, and four external calibration points (Table S5, S6).

Found at: doi:10.1371/journal.pone.0005615.s007 (5.76 MB TIF)

**Figure S2** Chronogram of salamander families calculated in r8s based on Bayesian analysis of *Rag1*, fixing the basal node at 250 MYA, and four external calibration points (Table S5, S6).

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## Author Contributions

Conceived and designed the experiments: RMB. Performed the experiments: RMB. Analyzed the data: RMB. Contributed reagents/materials/analysis tools: RMB PCC PEM RWVD DW. Wrote the paper: RMB. Contributed to manuscript preparation: PCC PEM RWVD DW. Contributed to funding: PCC DW.

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<b>Tree code/ map number</b>	<b>Locality State: County, location</b>	<b>Museum/ Field Number</b>	<b>Genbank Accession 16s</b>	<b>Genbank Accession Cytb</b>	<b>Genbank Accession Rag1</b>
<b><u>Amphiuma means (two-toed amphiuma):</u></b>					
1	AL: Covington, Pond Creek Bridge	AUM 37420	FJ951240	FJ951304	FJ951370
2	AL: Escambia, Conecuh N.F.	AUM 37447	FJ951241	FJ951305	FJ951371
3	FL: Alachua, River Styx	ASU 24181	FJ951242	FJ951306	FJ951372
4	FL: Alachua, Gainesville	RMB 3005	FJ951243	FJ951307	FJ951373
5	FL: Alachua, Orange Lake	MVZ 144889	FJ951244	FJ951308	-
6	FL: Baker, Ocoola N.F. Forest Rd. 241	MVZ 241516	FJ951245	FJ951309	-
7	FL: Glades, Pollywog Creek	UF 135378	FJ951246	FJ951310	FJ951374
8	FL: Leon, Clear Lake	MVZ 241517	FJ951247	FJ951311	-
9	FL: Leon, Silver Lake	MVZ 241518	FJ951248	FJ951312	-
10	FL: Levy, Gulf Hammock	APPSU 23768	FJ951249	AY691722	AY650127
11	FL: Liberty, Ochlockonee River	RMB 2983	FJ951250	FJ951313	FJ951375
12	FL: Okaloosa, Walton Pond	ASU 24552	FJ951251	FJ951314	FJ951376
13	FL: Orange, Shingle Creek	RMB 3006	FJ951252	FJ951315	FJ951377
14	FL: Putnam, Dunns Creek State Park	UF133757	FJ951253	FJ951316	FJ951378
15	FL: Putnam, Rodman Reservoir	RMB 3007	FJ951254	FJ951317	FJ951379
16	FL: Santa Rosa, Escambia River	ASU 25020	FJ951255	FJ951318	FJ951380
17	FL: Santa Rosa, near Munson	ASU 24554	FJ951256	FJ951319	FJ951381
18	FL: Taylor, US 27, 4 mi S of Madison Co	RMB 3001	FJ951257	FJ951320	FJ951382
19	GA: Liberty, near Hinesville	RMB 3131	FJ951258	FJ951321	FJ951383
20	GA: Liberty, near Hinesville	RMB 3132	FJ951259	FJ951322	FJ951384
21	GA: Liberty, near Hinesville	RMB 3133	FJ951260	FJ951323	FJ951385
22	LA: St. Tammany, Talisheek Bay	LSUMZ H-3333	FJ951261	FJ951324	FJ951386
23	LA: East Baton Rouge, Camp Istrouma	LSUMZ H-15033	FJ951262	FJ951325	FJ951387
24	NC: Columbus, Lake Waccamaw	RMB 3251	FJ951263	FJ951326	FJ951388
25	NC: Johnston, Clayton	MVZ 137236	FJ951264	FJ951327	-
26	NC: Johnston, Clayton	MVZ 137237	FJ951265	FJ951328	-
27	NC: Nash, near Stenhope	RMB 3187	FJ951266	FJ951329	FJ951389
28	NC: Wake, near Garner	RMB 3134	FJ951267	FJ951330	FJ951390
29	NC: Wake, near Garner	RMB 3135	FJ951268	FJ951331	FJ951391
30	SC: Berkeley, Francis Marion NF	RMB 3254	FJ951269	FJ951332	FJ951392
31	SC: Berkeley, Francis Marion NF	ASU 24509	FJ951270	FJ951333	FJ951393
32	VA: Charles City, Charles City	RMB 3249	FJ951271	FJ951334	FJ951394
33	VA: Charles City, Charles City	RMB 3250	FJ951272	FJ951335	FJ951395
<b><u>Amphiuma pholeter (one-toed amphiuma):</u></b>					
34	AL: Covington, Pond Creek	AUM 37412	FJ951273	FJ951336	FJ951396
35	FL: Hernando, Chassahowitzka WMA	RMB 3010	FJ951274	FJ951337	FJ951397
36	FL: Jefferson, near Capps	ASU 23947	FJ951275	FJ951338	-
37	FL: Jefferson, near Capps	ASU 23948	FJ951276	FJ951339	-
38	FL: Levy, Gulf Hammock	RMB 3255	FJ951277	FJ951340	FJ951398
39	FL: Levy, Gulf Hammock	APPSU 23767	FJ951278	AY691766	AY650128
40	FL: Walton, near Freeport	ASU 24330	FJ951279	FJ951341	FJ951399
41	FL: Walton, near Freeport	ASU 23914	FJ951280	FJ951342	FJ951400
42	GA: Grady, Ochlockonee River	RMB 3256	FJ951281	FJ951343	FJ951401
43	GA: Grady, Ochlockonee River	RMB 3257	FJ951282	FJ951344	FJ951402
44	GA: Grady, Ochlockonee River	RMB 3258	FJ951283	FJ951345	FJ951403
45	GA: Grady, Ochlockonee River	RMB 3259	FJ951284	FJ951346	FJ951404

**Amphiuma tridactylum (three-toed amphiuma):**

46	AR: Desha, T10S, R2W, S25	ASUMZ 29092	FJ951285	FJ951347	FJ951405
47	AR: Desha, T10S, R2W, S25	ASUMZ 29093	FJ951286	FJ951348	FJ951406
48	LA: Iberville, near Gabriel	LSUMZ H-877	FJ951287	FJ951349	FJ951407
49	LA: Livingston, ~2 mi NE of Watson	LSUMZ H-2793	FJ951288	FJ951350	FJ951408
50	LA: St. Landry, I-49, ~2 mi W of Whiteville	RMB 3093	FJ951289	FJ951351	FJ951409
51	LA: St. Martin, St. Martinville	BRM 1055	FJ951290	FJ951352	FJ951410
52	LA: St. Martin, St. Martinville	BRM 1056	FJ951291	FJ951353	FJ951411
53	LA: St. Tammany, Talisheek Bay	LSUMZ H-3334	FJ951292	FJ951354	FJ951412
54	LA: Vermillion, ~5 km NW of Andrew	BRM 1075	FJ951293	FJ951355	FJ951413
55	LA: Vermillion, ~5 km NW of Andrew	BRM 1076	FJ951294	FJ951355	FJ951414
56	LA: Vernon, Kisatchie NF	MVZ 241480	FJ951295	FJ951356	FJ951415
57	MS: Pearl River	ASU 10218	FJ951296	FJ951357	FJ951416
58	TN: Lake or Obion, Reel Foot Lake	RMB 3136	FJ951297	FJ951358	FJ951417
59	TX: Jefferson, Port Arthur	RMB 3253	FJ951298	FJ951359	FJ951369
60	TX: Jefferson, Port Arthur	RMB 2982	FJ951299	FJ951360	FJ951418

**Outgroups:**

Plethodontidae: <i>Plethodon cinereus</i>	AY728232	AY728232	AY691745
Rhyacotritonidae: <i>Rhyacotriton variegatus</i>	AY728219	AY728219	AY691693
Ambystomatidae: <i>Ambystoma mexicanum</i>	AY659991	AY659991	EF107293

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APPSU = Appalachian State University

ASUMZ = Arkansas State University Museum of Zoology

AUM = Auburn University Natural History Museum

BRM = Brad R. Moon field series

LSUMZ = Museum of Natural Science, Louisiana State University

MVZ = Museum of Vertebrate Zoology, University of California, Berkeley

RMB = Ronald M. Bonett field series

UF = Florida Museum of Natural History, University of Florida

UTA = Reptile Diversity Research Center, University of Texas at Arlington

- = no sequence collected for the individual

<b>Family:</b>	<b>Genbank Accession</b>	<b>Genbank Accession</b>	<b>Genbank Accession</b>	<b>Genbank Accession</b>	<b>Genbank Accession</b>	<b>Genbank Accession</b>
<b>Species</b>	<i>Pomc</i>	<i>Ncx1</i>	<i>Slc8a3</i>	<i>ND2, &amp; tRNAs</i>	<i>Cox1</i>	<i>Nd4 &amp; tRNAs</i>
<b><u>Ambystomatidae:</u></b>						
<i>Ambystoma mexicanum</i>	EU275841*	EF107230	EF107367	AY659991	AY659991	AY659991
<b><u>Rhyacotritonidae:</u></b>						
<i>Rhyacotriton variegatus</i>	EU275823	EF107242	EF107401	AY728219	AY728219	AY728219
<b><u>Plethodontidae:</u></b>						
<i>Plethodon cinereus</i>	FJ951365	EF107226	EF107360	AY728232	AY728232	AY728232
<b><u>Amphiumidae:</u></b>						
<i>Amphiuma means</i>	FJ951366	FJ951362	FJ951420	AY916037	FJ951301	AY691766
<i>Amphiuma pholeter</i>	FJ951367	FJ951363	FJ951421	AY916035	FJ951302	AY691767
<i>Amphiuma tridactylum</i>	FJ951368	FJ951361	FJ951419	AY916036	FJ951300	FJ951364

\* *Ambystoma rosaceum*

<b>Gene</b>	<b>Primer Name</b>	<b>Primer sequence</b>	<b>Size* (bp)</b>	<b>Reference</b>
<i>16s</i>	16Sa-L	5'-CGCCTGTTTATCAAAAACAT-3'	538	[32]
	16Sb-H	5'-CCGGTCTGAACTCAGATCACGT-3'		
<i>Cytb</i>	MVZ15	5'-GAACTAATGGCCACACWWTACGNAA-3'	783	[33]
	MVZ16	5'-AAATAGGAARTATCAYTCTGGTTTRAT-3'		
<i>Nd4</i>	ND4F	5'-CACCTATGACTACCAAAGCTCATGTAGAAGC-3'	885	[34]
	LeuR	5'-CATTACTTTTACTTGGATTGACCA-3'		
<i>Col</i>	MVZ_201	5'-TCAACAAAYCATAAAGATATTGGCACC-3'	1260	[35]
	MVZ_202	5'-GCGTCWGGGTARTCTGAATATCGTCG-3'		
<i>Rag1</i>	AmphRag1F	5'-TGCCCGGCTATCATCCCTTTGAAT-3'	825	This study
	AmphRag1R	5'-ACCTCCCTCACAAGCTTCTCATCA-3'		
<i>Pomc</i>	POMC_Amphiu_F	5'-ATATGTCATGAGCCATTTTCGCTGGAA-3'	481	This study
	POMC_Amphiu_R	5'-GGCATTTTTGAAAAGAGTCATTAGAGG-3'		
<i>Ncx1</i>	NCX1_Amphiu_A_F	5'-GCACTATTGTTGGAAGTGCTGCCT-3'	814	This study
	NCX1_Amphiu_A_R	5'-TCAGATCCTGCATTAGCTGTGCCA-3'		
<i>Slc8a3</i>	SLC8A3_Amphiu_A_F	5'-TGCGGTCATGGGTTTATAGCTGGA-3'	761	This study
	SLC8A3_Amphiu_A_R	5'-ATGTCACCACCCTTACGAACCACA-3'		

Ambiguity codes: N = G, A, T or C; R = A or G; W = A or T; Y = C or T.

\* Fragment size between the primer pairs for each gene.

Gene/partition	# of positions			
	in partition	nst	rates	statefreqpr
<i>Rag1</i> : alignment pos 1, codon pos 1	492	6	gamma	dirichlet (1,1,1,1)
<i>Rag1</i> : alignment pos 2, codon pos 2	491	1	equal	dirichlet (1,1,1,1)
<i>Rag1</i> : alignment pos 3, codon pos 3	491	6	gamma	dirichlet (1,1,1,1)
<i>Pomc</i> : alignment pos 1, codon pos 3	161	6	gamma	dirichlet (1,1,1,1)
<i>Pomc</i> : alignment pos 2, codon pos 1	160	2	equal	dirichlet (1,1,1,1)
<i>Pomc</i> : alignment pos 3, codon pos 2	160	6	equal	dirichlet (1,1,1,1)
<i>Ncx1</i> : alignment pos 1, codon pos 3	245	6	equal	fixed (equal)
<i>Ncx1</i> : alignment pos 2, codon pos 1	245	6	propinv	dirichlet (1,1,1,1)
<i>Ncx1</i> : alignment pos 3, codon pos 2	245	1	equal	dirichlet (1,1,1,1)
<i>Slc8a3</i> : alignment pos 1, codon pos 2	86	1	equal	dirichlet (1,1,1,1)
<i>Slc8a3</i> : alignment pos 2, codon pos 3	86	2	equal	dirichlet (1,1,1,1)
<i>Slc8a3</i> : alignment pos 3, codon pos 1	86	1	equal	dirichlet (1,1,1,1)
<i>16s</i>	377	6	propinv	dirichlet (1,1,1,1)
<i>Nad1</i> : alignment pos 1, codon pos 1	112	2	propinv	dirichlet (1,1,1,1)
<i>Nad1</i> : alignment pos 2, codon pos 2	112	2	propinv	dirichlet (1,1,1,1)
<i>Nad1</i> : alignment pos 3, codon pos 3	111	2	equal	dirichlet (1,1,1,1)
<i>Nad2</i> : alignment pos 1, codon pos 1	337	6	gamma	dirichlet (1,1,1,1)
<i>Nad2</i> : alignment pos 2, codon pos 2	337	6	propinv	dirichlet (1,1,1,1)
<i>Nad2</i> : alignment pos 3, codon pos 3	336	6	propinv	dirichlet (1,1,1,1)
<i>Co1</i> : alignment pos 1, codon pos 1	173	6	gamma	fixed (equal)
<i>Co1</i> : alignment pos 2, codon pos 2	172	1	equal	dirichlet (1,1,1,1)
<i>Co1</i> : alignment pos 3, codon pos 3	172	6	gamma	dirichlet (1,1,1,1)
<i>tRNAs: His, Ile, Met, Trp, Ala, Asn, Tyr</i>	421	6	gamma	dirichlet (1,1,1,1)
<i>Nad4</i> : alignment pos 1, codon pos 1	210	6	gamma	dirichlet (1,1,1,1)
<i>Nad4</i> : alignment pos 2, codon pos 2	210	2	propinv	dirichlet (1,1,1,1)
<i>Nad4</i> : alignment pos 3, codon pos 3	209	2	propinv	dirichlet (1,1,1,1)
<i>Cytb</i> : alignment pos 1, codon pos 2	217	1	equal	dirichlet (1,1,1,1)
<i>Cytb</i> : alignment pos 2, codon pos 3	217	2	equal	dirichlet (1,1,1,1)
<i>Cytb</i> : alignment pos 3, codon pos 1	217	1	equal	dirichlet (1,1,1,1)

<b>Family</b>	<b>Species</b>	<b>Genbank Accession Rag1</b>
Ichthyophiidae (Caecilian)	<i>Ichthyophis</i> sp.	AY650147
Ascaphidae (Anuran)	<i>Ascaphus montanus</i>	AY650146
Cryptobranchidae	<i>Andrias davidianus</i>	AY650142
Salamandridae	<i>Salamandra salamandra</i>	AY650135
Salamandridae	<i>Taricha rivularis</i>	AY650133
Ambystomatidae	<i>Ambystoma mexicanum</i>	EF551561
Dicamptodontidae	<i>Dicamptodon tenebrosus</i>	AY650132
Rhyacotritonidae	<i>Rhyacotriton variagatus</i>	AY691693
Amphiumidae	<i>Amphiuma means</i>	AY650127
Amphiumidae	<i>Amphiuma pholeter</i>	AY650128
Amphiumidae	<i>Amphiuma tridactylum</i>	FJ951369
Plethodontidae	<i>Aneides aeneus</i>	AY691701
Plethodontidae	<i>Aneides lugubris</i>	AY650118
Plethodontidae	<i>Bolitoglossa helmrichi</i>	AY650124
Plethodontidae	<i>Desmognathus brimleyorum</i>	AY691697
Plethodontidae	<i>Ensatina eschscholtzii</i>	AY691743
Plethodontidae	<i>Eurycea bislineata</i>	AY691706
Plethodontidae	<i>Hemidactylium scutatum</i>	AY691711
Plethodontidae	<i>Phaeognathus hubrichti</i>	AY691700
Plethodontidae	<i>Plethodon cinereus</i>	AY691703

\*All tree partitions were set to: nst=6, rates=invgamma, statefreqpr=dirichlet (1,1,1,1).

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#- Fossil and/or description of calibration point	Date	Reference
1* - <i>Chunerpeton tianyiensis</i> , fixed MRCA of cryptobranchids and salamandroids	161 MYA	[46]
2*- Estimated molecular divergence of cryptobranchids and salamandroids.	250 MYA	[30, 31, 47]
3 - <i>Proamphiuma cretacea</i> , minimum MRCA of amphiumids and plethodontids	65.5 MYA	[48, 49]
4 - <i>Dicamptodon antiquus</i> , minimum MRCA of dicamptodontids and ambystomatids	58 MYA	[50]
5- <i>Aneides</i> sp., minimum MRCA of <i>Aneides</i> and desmognathines	19 MYA	[51]
6 - <i>Aneides lugubris</i> , minimum MRCA <i>Aneides</i> from eastern and western North America	5 MYA	[52]

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MRCA = Most recent common ancestor

MYA = Million years ago

\*Note: analyses are based on either fixing the root node (MRCA of cryptobranchids and salamandroids) at either 161 MYA (1) or 250 MYA (2). See methods section.

